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Gaussian-Based Dilated 1-D CNN for Classifying B-Cell Epitopes in Zika and Dengue Protein Sequences



Abstract: - The main goal of designing peptide vaccines, conducting immunodiagnosis, and producing antibodies is to accurately identify linear B-cell epitopes. However, experimental analysis to determine these epitopes is costly. This study focuses on developing a Gaussian-based dilated 1-D CNN model for classifying epitopes and non-epitopes in protein sequences related to Zika and Dengue viruses. The Immune Epitope Database (IEDB) was used, containing a total of 1741 and 7020 linear B-cell epitopes for Zika and Dengue viruses, respectively. Physicochemical features of the protein sequences dataset were extracted using the Gaussian distribution to extract optimal features based on feature probability distribution. The proposed model achieved an accuracy score of 83.00% and 85.00%, precision of 87.00%, recall of 83.00% and 85.00%, and an F1-score of 84.00% and 86.00% over the Zika and Dengue datasets. The suggested model outperforms existing methods, demonstrating the potential of deep learning approaches in bioinformatics for enhancing epitope prediction in viruses, with implications for drug discovery and vaccine development.

Keywords: Linear B-Cell, Protein Sequences, Amino Acid Sequences, Epitopes, Non-Epitopes, Deep Learning, Drug Discovery, Vaccine Development

I. INTRODUCTION

The human immune system's reaction relies heavily on antibodies, which are essential elements that identify and attach to the proteins of pathogenic organisms like bacteria or viruses [1,51,52,53,54]. An epitope represents the portion of an antigenic material that such antibodies identify. It is possible to identify a linear epitope, that is an ongoing chain of amino acids found within the linear protein sequence, as well as a conformational epitope, which is a group of amino acids that may be divided in the amino acid sequence but are situated strongly in the three-dimensional framework of the protein. In instances, uses like peptide-based vaccine development [2,55,56,57,58], immuno-diagnostic testing [3], and the synthesis of synthesized antibodies [47,48,49,50] depend on the recognition B-cell epitopes (BCEs). Statistical modeling can be crucial in the invention of novel vaccines and medications toward major viruses infections such as the HIV, liver disease, or flu viruses, since clinical identification

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of BCEs is costly and time-consuming [5]-[6]-[7]. The forecasting of continuous BCEs has drawn a lot of interest [8], despite the fact that conformational BCEs make up almost all of normally existing BCEs [9]. This is because linear BCEs are useful for peptide-based vaccine production, besides additional uses [10]-[11].

Some physicochemical feature of the individual amino acids, such as membrane access [12], fluidity [13], hydrophilic properties [14], or antigenic properties [15], was the single assessed by the early epitope predicting algorithms. Among the techniques which are presently available online include BEPIPOPE [16], PEOPLE [17]. Using a folding frame across the search query peptide sequence, these methods determine the typical amino acid probability score for each characteristic [37,38,39,40]. A linear BCE is identified in the corresponding area of the sequence whenever the projected ratios for an ongoing portion of the amino acid are higher than a predetermined cut-off. On the other hand, using a particular amino acid profile or additionally a mixture of traits, an evaluation of 485 likelihood factors showed that these factors are ineffective to identify BCEs and slightly exceeded randomized BCE identification [41,42,43].

Innovative methods that were based on multiple likelihood factors and incorporated previously unincluded amino acid characteristics have been developed in response to the growing accessibility of empirically determined epitopes [19]. These techniques, which differentiate between BCEs and non-BCEs in the arrangement of amino acids using machine learning (ML) techniques, have demonstrated higher performance than individual likelihood scale-based techniques. BCEs are provided as features sets for learning the machine learning models, which originate from various features of amino acids, including the amino acid composition (AAC), the amino acid pair antigenicity level [20]. BepiPred 3.0 [21], ABCPred [22], AAAPred [23], SVMTrip [24], EpitopeVec [18], and EpiDope [25], EpitopeVec [26] represent a few instances of ML-based techniques for BCE modeling. One prominent problem appears to be that none of the previously listed techniques achieve higher performance when used in a cross-testing strategy, because ML training and validation are carried out on separate databases [44,45,46].

Contribution of the paper

- To develop the Gaussian based dilated 1-D CNN for classifying epitopes and non-epitopes in protein sequences associated with Zika and Dengue viruses.
- This study's contribution lies in its application of deep learning techniques to bioinformatics, aiming to improve epitope prediction in viruses. Such models have the potential to enhance for understanding of viral proteins, aiding in drug discovery and vaccine development efforts. The research underscores the importance of deep learning in bioinformatics and its potential to impact public health and medical research positively.

Organization of the paper

Section 2 presents the brief overview of dataset, Physicochemical feature extraction, Gaussian Distribution function, deep learning model. The suggested methodology and dilated 1D-CNN model are presented in section 3. Section 4 presents the result analysis, comparative analysis of the proposed model. Section 5 discussed the conclusion and future direction of the study.

II. MATERIALS AND METHODS

Figure 1 shows the architecture of proposed model which consists of data inputs, feature extraction, building Deep learning model, and classification. Input of the model are protein sequence of Zika and Dengue virus dataset that consists of 20 amino acid characters. The Physicochemical Features of 21-character text sequences are provided as inputs. These features need to be converted into an integer sequence using quantization coding. These features are extracted using Gaussian distribution to extract the optimal features based on the probability of feature distribution. The dilated 1D-CNN model is designed for classifying the epitopes and non-epitopes of protein sequences.

Datasets

In this study, extracted the peptide dataset of Zika and Dengue from the IEDB that contains the epitopes and non-epitopes. (<http://www.iedb.org/>). The IEDB database recorded a total of 1741 and 7020 linear B-cell epitopes of zika and dengue respectively. Among these total epitopes, 1261 positive epitopes and 480 negative epitopes were

recorded from the Zika virus dataset. Likewise, 5008 positive epitopes and 2012 negative epitopes are recorded from the dengue virus dataset.

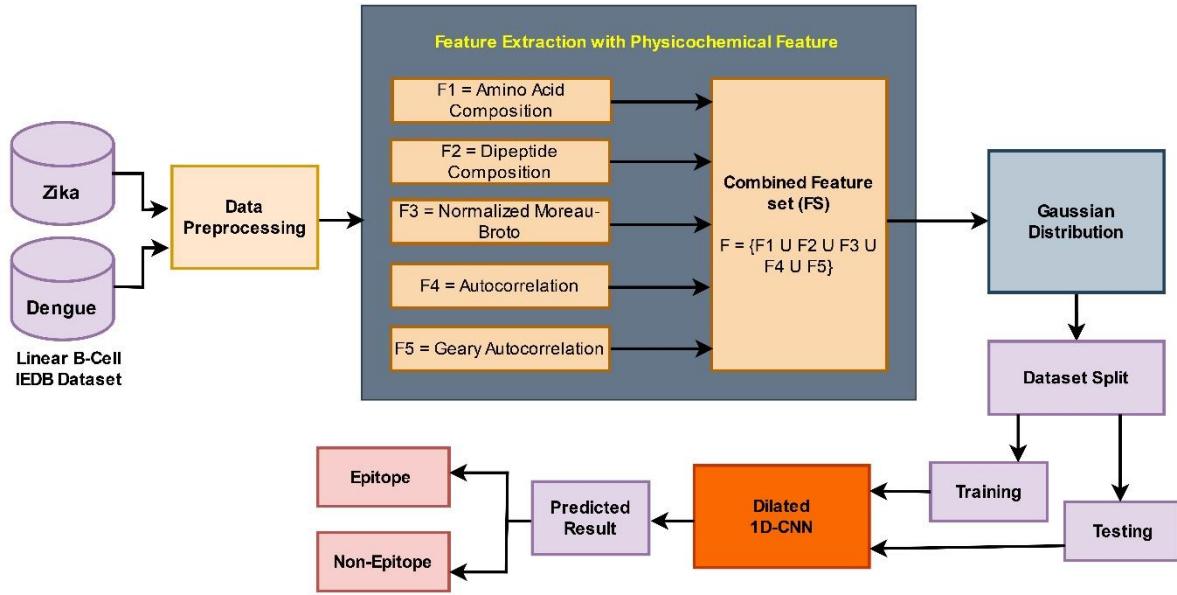


Figure 1: Architecture of Proposed Model

Physicochemical Feature

Amino Acid Composition : A vector indicating the proportional amount of every single amino acid in the protein serves as a representation of the AAC [27]. It could be expressed as:

$$ACC = (f1, f2, \dots, f20) \quad (1)$$

Where $f_i = \frac{A_i}{N}$ ($i=1,2,3,\dots,20$) shows the type of amino acid i , A_i represents the total amount of amino acid sequence, and N represents the length of amino acid sequence

Dipeptide Composition

A vector that specifies the quantity of dipeptides standardized over every possible dipeptide pairings for a given protein sequence is used to express dipeptide composition (DC). Its features remain constant at 400 in length [28]. It could be expressed as:

$$DC = (f1, f2, \dots, f400) \quad (2)$$

Where $f_i = \frac{A_i}{N}$ ($i=1,2,3,\dots,400$) shows the type of dipeptide i , A_i represents the total amount of dipeptide composition, and N represents the length of peptide

Moreau-Broto Autocorrelation

The Moreau-Broto autocorrelation feature is a method used to analyze protein sequences. It calculates the correlation between the properties of amino acids in a protein sequence and their positions within the sequence. Mathematically, the Moreau-Broto autocorrelation for a feature P of an amino acid sequence is given by:

$$MB-autocorrelation(P) = \sum_{i=1}^{n-1} \left(\frac{P(i) \cdot P(i+1)}{i} \right) \quad (3)$$

where n is the length of the sequence, and $P(i)$ represents the value of property P for the amino acid at position i in the sequence. This feature helps in predicting various properties of proteins, such as their function, with other molecules, by considering the spatial relationships between amino acids in the sequence.

Moran Autocorrelation

The Moran autocorrelation feature is a method used in bioinformatics to analyze protein sequences [29]. It calculates the correlation between the properties of amino acids in a protein sequence and the properties of neighboring amino acids. Mathematically, the Moran autocorrelation for a property P of an amino acid sequence is given by:

$$\text{Moran - autocorrelation}(P) = \frac{n}{\sum_{i=1}^n (P(i) - \bar{P})^2} \cdot \frac{\sum_{i=1}^n \sum_{j=i+1}^n (P(i) - \bar{P})(P(j) - \bar{P})}{\sum_{i=1}^n (P(i) - \bar{P})^2} \quad (4)$$

Where n is the length of the sequence, $P(i)$ represents the value of property P for the amino acid at position i in the sequence, and \bar{P} is the average value of property P across all amino acids in the sequence. This feature helps in predicting various properties of proteins by considering the spatial relationships between amino acids and their properties.

Geary Autocorrelation

The Geary autocorrelation feature is a method used to measures the similarity between the properties of amino acids at different positions in the sequence. Mathematically, the Geary autocorrelation for a property P of an amino acid sequence is given by:

$$\text{Geary - autocorrelation}(P) = \frac{\sum_{i=1}^n \sum_{j=i+1}^n (P(i) - P(j))^2 \cdot w_{ij}}{2 \cdot \sum_{i=1}^n (P(i) - P(j))^2} \quad (5)$$

Where n is the length of the sequence, $P(i)$ represents the value of property P for the amino acid at position i in the sequence, and w_{ij} is a weight factor that can be defined based on the distance between positions i and j in the sequence. This feature helps in predicting various properties of proteins by considering the spatial relationships between amino acids and their properties.

Combined Feature

In this step combine the features can enhance the prediction of model by incorporating diverse information of amino acid sequences over the Zika and Dengue dataset. In this step combining all five physicochemical properties of amino acid sequence that can lead to improved accuracy in predicting epitopes.

$$F = ACC \cup DC \cup MBA \cup MA \cup GA \quad (6)$$

Gaussian Distribution

In this study, presented the Gaussian distribution to extract the optimal features for enhancing the weights of the targeted features and its adjacent features [30], so that the proposed deep model can train with optimal features of amino acid. The Gaussian distribution function is:

$$f(a) = \frac{1}{\sqrt{2\pi\delta}} \exp\left(-\frac{(a-\mu)^2}{2\delta^2}\right) \quad (7)$$

Gaussian distribution is measured using equations 8

$$f(a) = \int_{-\infty}^a f(a) dx, \quad (8)$$

Gaussian probability distribution function is defined in equation 9

$$P(a) = F(a) - F(x - w) \quad (9)$$

where w represents the token window, δ is the standard deviation, μ is the average of the distribution, and a is an actual value. To denote the length of every token in the tests, defined the token window w to 1. It is also defined the optimum values of μ and δ to 0 and 2.5, respectively. The design set is where these properties are adjusted. This probability is used as a feature in deep learning model for protein sequence prediction.

Dilated 1D-CNN

In this study, we designed a dilated 1-D CNN to classify epitopes and non-epitopes from protein sequences related to Zika and Dengue viruses. Before designing the proposed dilated 1D-CNN model, we set the parameters for training the model, which includes 3 channels with an input width of 17, 32 filters, a kernel size of 3, and a

dilation rate of 2. The model was trained with a batch size of 32 over 100 epochs using the Adam optimizer. The model's loss was measured using the binary cross-entropy function.

Given 1D protein sequence features $f: N \rightarrow R$ and kernel $k: \{0, 1, \dots, n-1\} \rightarrow R$, the dilated convolutional function $(f * d k): N \rightarrow R$ is:

$$(f * d k)(s) = \sum_{i=0}^{n-1} k(i) * (s - id) \quad (10)$$

Where, N is the real numbers, and n and d represents the kernel size and dilation parameter respectively. If $d = 1$, the neural network operate normal convoluptonal operator. In dilated CNN, residual connection is used to stability of the network [31]. In this network used 1×1 convolutional layer to compare the size of input and output. The weight normalization is used in the kernel of dilated layer. To randomized apply the dropout to the output layer. The Relu is used as a activation function . The figure 2 shows the complete architecture of the 1D dialted CNN model.

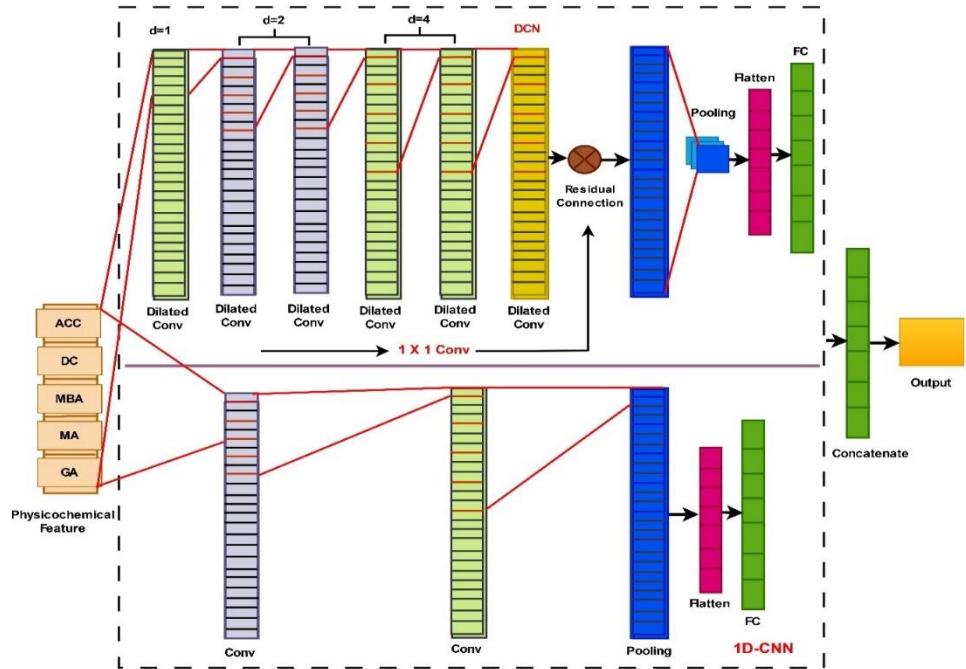


Figure 2: Architecture of the Dilated 1D CNN Model

Dilated Convolutional Operator

The dilated convolutional function can be defined $(F *_d k): N \rightarrow C$ as follows

$$(F *_d k)(s) = \arg \min_C \sum_{i=0}^{N-1} k(i) d_C^2(F(s - id), C) \quad (11)$$

Where, C represents the valued operator of $(F *_d k)$. the C is used as manifold function which is equal to Euclidean dilated layer.

Residual Connection

If F and Y is the input and ouput of neural network. Based on the Euclidean residual connection, apply residual connection in two phases: 1) Concatenate F and $Y(F)$ to obtained the number of input and output channels. 2) wYC is used extract the ouput. Let, $R(F, Y_f)$ is the ouput of the residual connection [32], then n^{th} channel of connection , $R_n(F, Y(F))$ is defined as

$$R_n(F, Y(F))(s) \stackrel{\text{def}}{=} \arg \min_C \quad (12)$$

$$\left(\sum_{i=1}^{input} k(i) d_C^2(F(s - id), C) + \sum_{j=1}^{count} k(j + input) d_C^2(Y_j(s), C) \right),$$

$$St \sum_i k(i) = 1, \forall k(i) > 0 \quad (13)$$

Where, $n \in (1, 2, \dots, \text{output})$ and F_i and Y_j represents the i^{th} and j^{th} channel of F and Y respectively.

Loss Function

The binary cross-entropy loss function is used for classifying epitopes and non-epitopes. It calculates the difference between the predicted probability distribution and the actual distribution of the epitopes and non-epitopes. Mathematically, it is defined as:

$$\text{Binary Cross - Entropy Loss} = -\frac{1}{N} \sum_{i=1}^N [y_i \log(\hat{y}_i) + (1 - y_i) \log(1 - \hat{y}_i)] \quad (14)$$

Where N is the number of features, y_i is the actual label (epitopes and non-epitopes) for features i , and \hat{y}_i is the predicted probability of the features belonging to epitopes. The loss function penalizes the model more when it makes incorrect predictions with high confidence and less when it is uncertain. The goal is to minimize this loss function during training to improve the model's ability to correctly classify epitopes and non-epitopes.

Pseudo Code: Dilated 1D-CNN

Input: $F = F(\text{ACC}, \text{DC}, \text{MBA}, \text{MA}, \text{GA})$ // Physicochemical Feature

Output: *Classify Epitope and Non - Epitope*

Define the Gaussian distribution function based on equation 6

Measure the Gaussian distribution using the cumulative distribution function based on equation 7

Define the Gaussian probability distribution function based on equation 8

Function 1D-CNN

Parameters ($N, F_{in}, F_{out}, \text{res}, k1, d1, k2, d2, nC, c$)

$x^{i-1} = \text{Input}(F_{in}, N)$

$y1 = \text{Dilated - Conv}(x^{i-1}, F_{in}, F_{out}, k1, d1)$

$y1 = \text{Dilated - Conv}(y1, F_{out}, k2, d2)$

$x^i = \text{Residual}(x^{i-1}, y1, F_{in}, F_{out}, C_{res})$

$y0 = \text{Inv}(x^i, nC, c)$

End Function

The above pseudo code describes a dilated 1D-CNN for classifying epitope and non-epitope regions in protein sequences based on physicochemical features (ACC, DC, MBA, MA, GA). It begins by defining a Gaussian distribution function and measuring it using a cumulative distribution function. The 1D-CNN function takes parameters such as the number of amino acids (N), input and output features (F_{in}, F_{out}), kernel sizes ($k1, k2$), dilation rates ($d1, d2$), and number of classes (nC). It then performs dilated convolutions with different kernel sizes and dilation rates, calculates the residual connection between the input and output, and performs an inverse transformation to obtain the classification probabilities for epitope and non-epitope regions.

III. RESULT ANALYSIS

The proposed Gaussian based Dilated 1D-CNN model was trained using Physicochemical Features of linear B-Cell protein sequence over the Zika and Dengue virus dataset. The implementation was carried out using core Python programming and the scikit-learn library. The experimental setup was conducted on Google Colab, utilizing a high-end GPU and 16 GB of RAM. To measure the performance and effectiveness of proposed Gaussian based Dilated 1D-CNN model for prediction and classification of linear B-cell Epitopes. Following evaluation parameters such as accuracy, precision, Recall and f1 score is used to meets the desired objectives of the study.

$$\text{Accuracy} = \frac{T_P + T_N}{T_P + T_N + F_P + F_N} \quad (15)$$

$$\text{Precision} = \frac{T_P}{T_P + F_P} \quad (16)$$

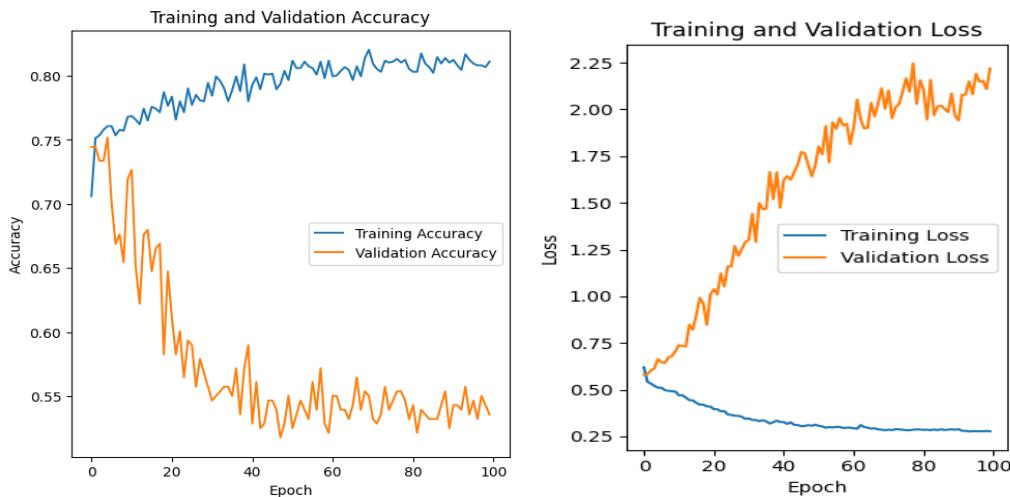
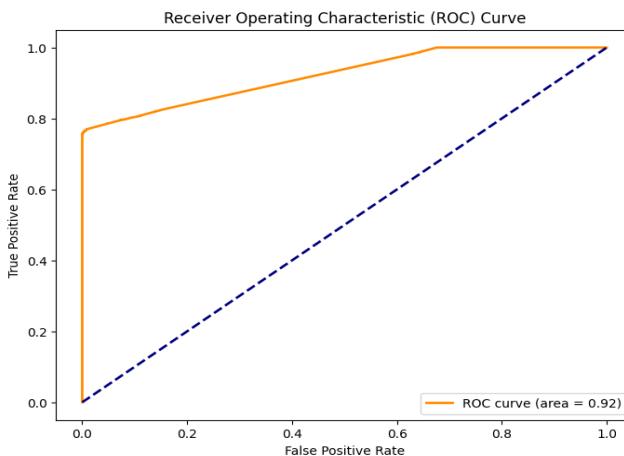
$$\text{Recall} = \frac{T_P}{T_P + F_N} \quad (17)$$

$$\text{F1 - Score} = 2 \times \frac{\text{Precision} \times \text{Recall}}{\text{Precision} + \text{Recall}} \quad (18)$$

Table 1: Classification Report of proposed model over the Zika virus dataset

	Precision	Recall	F1-Score	Support
0	0.65	0.90	0.75	395
1	0.95	0.80	0.87	997
Accuracy			0.83	1392
Micro Avg	0.80	0.85	0.81	1392
Weighted Avg	0.87	0.83	0.84	1392

Table 1 shows the classification report of a proposed model for classifying epitope and non-epitope of linear B-cell over the Zika virus dataset. For class 0 (epitope), the model achieved a precision of 0.65, recall of 0.90, and F1-score of 0.75. For class 1 (non-epitope), the precision was higher at 0.95, recall was 0.80, and F1-score was 0.87. The accuracy of the model was 0.83. In the micro-average calculation across both classes, the precision was 0.80, recall was 0.85, and F1-score was 0.81, considering all instances. The weighted average, which considers class imbalance, resulted in a precision of 0.87, recall of 0.83, and F1-score of 0.84. This indicates that the model performed well, particularly in classifying class 1 instances.

**Figure 3: Training and Validation Accuracy and Loss of model for Zika virus dataset****Figure 4: ROC curve for Zika dataset**

The figure 3 shows the training and validation accuracy and loss of a model trained on the Zika virus dataset. The training accuracy steadily increases over 100 epochs. The training loss, which measures the difference between predicted and actual values during training, decreases over 100 epochs, indicating that the model is improving in its predictions. The figure 3 indicates that insights into how well the model is learning and generalizing from the

Zika data. Figure 4 shows the ROC curve that indicate the performance of a dilated 1D-CNN model trained on the Dengue virus dataset in terms of its true positive rate against the false positive rate across different thresholds. The area under the ROC curve (AUC) is 0.92, indicating that the model has good discriminative ability in distinguishing between positive and negative instances. A higher AUC value suggests that the model is better at classifying Epitopes and Non-Epitopes correctly across various thresholds. The ROC curve and its AUC of 0.92 indicate that the suggested model has a strong performance in classifying Epitopes and Non-Epitopes in the Zika virus dataset.

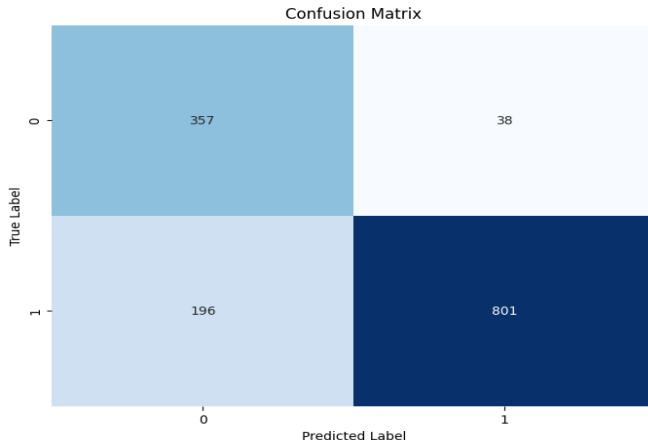


Figure 5: Confusion Matrix for Zika Dataset

Figure 5 shows the confusion matrix, indicates that the model correctly identified 196 epitope regions and 38 non-epitope regions, but it misclassified 801 non-epitope regions as epitope and 357 epitope regions as non-epitope over the Zika Dataset.

Table 2: Classification Report of Proposed Model over the Dengue virus dataset

	Precision	Recall	F1-Score	Support
0	0.70	0.86	0.77	1616
1	0.94	0.85	0.89	3999
Accuracy			0.85	5615
Micro Avg	0.82	0.85	0.83	5615
Weighted Avg	0.87	0.85	0.86	5615

Table 2 presents the classification report of a proposed model for classifying epitope and non-epitope of linear B-cell over the Dengue virus dataset. For class 0 (non-epitope), the model achieved a precision of 0.70, recall of 0.86, and F1-score of 0.77. For class 1 (non-epitope), the precision was higher at 0.94, recall was 0.85, and F1-score was 0.89. The overall accuracy of the model was 0.85.

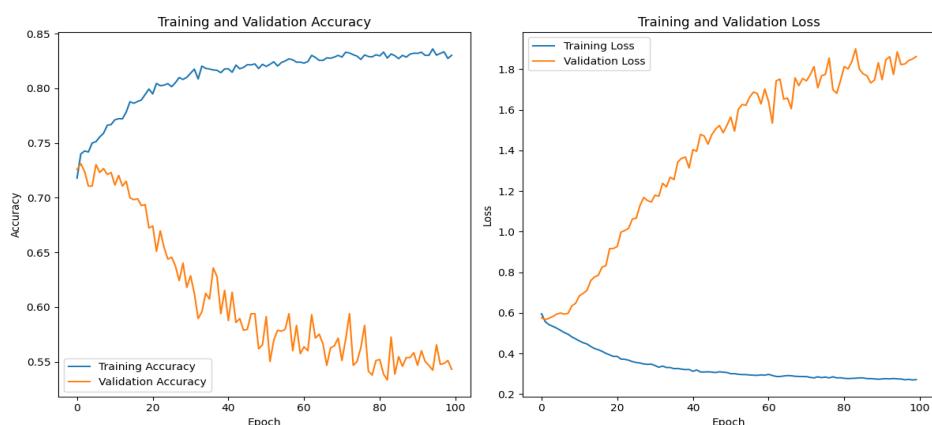


Figure 6: Training and Validation Accuracy and Loss of model for Dengue virus dataset

Figure 6 shows the training and validation accuracy and loss of a model trained on the Dengue virus dataset. The training accuracy steadily increases over 100 epochs. The training loss, which measures the difference between predicted and actual values during training, decreases over 100 epochs, indicating that the model is improving in its predictions. The figure 6 indicates that insights into how well the model is learning and generalizing from the Zika data.

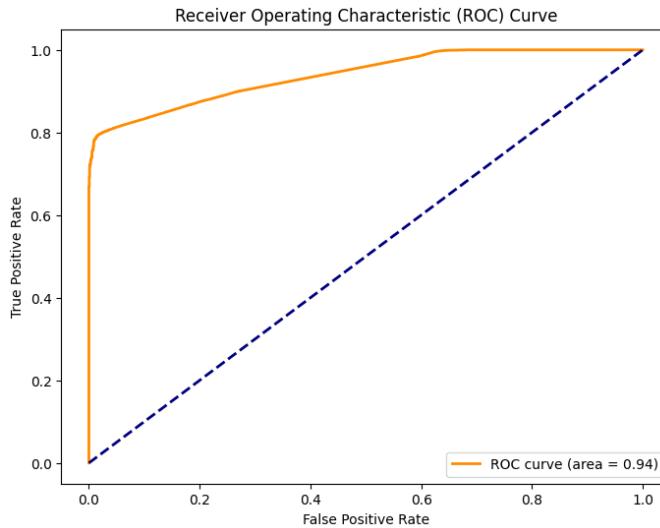


Figure 7: ROC Curve of Model for Dengue Dataset

Figure 7 shows the ROC curve that indicate the performance of a dilated 1D-CNN model trained on the Dengue virus dataset in terms of its true positive rate against the false positive rate across different thresholds. The area under the ROC curve (AUC) is 0.94, indicating that the model has good discriminative ability in distinguishing between positive and negative instances. A higher AUC value suggests that the model is better at classifying Epitopes and Non-Epitopes correctly across various thresholds. Overall, the ROC curve and its AUC of 0.94 indicate that the model has a strong performance in classifying Epitopes and Non-Epitopes in the Dengue virus dataset.

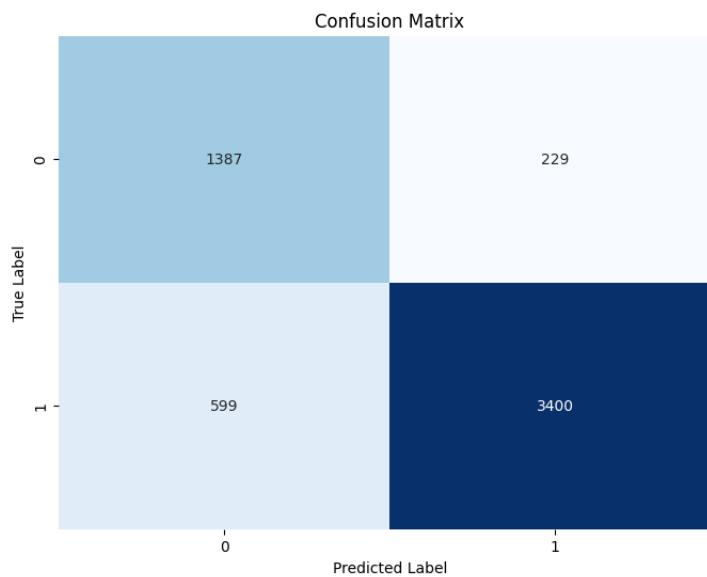


Figure 8: Confusion Matrix for Dengue Dataset

Figure 8 shows the confusion matrix, indicates that the model correctly identified 599 epitope regions and 229 non-epitope regions, but it misclassified 1387 non-epitope regions as epitope and 3400 epitope regions as non-epitope over the Dengue Dataset.

Table 3: Comparative Analysis of Proposed Model with Existing Methods

Author	Dataset	Accuracy	Precision	Recall	F1-Score
Liu F. et. al. (2024) [33]	iBCE-EL	75.7	72.2	73.1	72.6
Ras-Carmona A et. al. (2022) [34]	BLAST	72.54	81.58	63.49	-
Khanna D. et. al. (2020) [35]	APCpred	76.6	70.00	82.00	-
Maximilian Collatz et. al. (2020) [25]	Bepipred	69.00	63.5	52.00	-
Qi Y. et. al. (2023) [36]	DeepLBCEPred	67.00	63.00	71.00	-
Proposed Dilated 1D-CNN	Zika	83.00	87.00	83.00	84.00
	Dengue	85.00	87.00	85.00	86.00

Table 3 presents a comparative analysis of the proposed model with existing methods for classifying epitope and non-epitope. The proposed Dilated 1D-CNN model outperforms existing methods in terms of accuracy, precision, recall, and F1-score. The proposed model achieves an accuracy of 83%, which is higher than the accuracies reported by Liu F. et al. (75.7%), Ras-Carmona A et. al. (72.54%), Khanna D. et. al. (76.6%), Maximilian Collatz et. al. (69.00%), and Qi Y. et. al. (67.00%). Similarly, the precision, recall, and F1-score of the proposed model are also higher compared to the existing methods, indicating that the proposed Dilated 1D-CNN model is more effective in classifying epitope and non-epitope regions in the dataset.

IV. CONCLUSION AND FUTURE SCOPE

In this study, we addressed the challenge of accurately identifying linear B-cell epitopes in protein sequences related to Zika and Dengue viruses. We introduced a Gaussian-based dilated 1-D CNN model, aiming to improve epitope prediction, a critical step in peptide vaccine design and immunodiagnosis. Our model, trained on the Immune Epitope Database (IEDB) containing 1741 and 7020 epitopes for Zika and Dengue dataset, respectively, achieved high accuracy score of 83.00% and 85.00%, precision of 87.00%, recall of 83.00% and 85.00%, and an F1-score of 84.00% and 86.00% over the Zika and Dengue datasets. The results indicate the effectiveness of our model in classifying epitopes and non-epitopes, outperforming existing methods. For future research, we plan to enhance the model by exploring advanced deep learning pretrained techniques such as Resnet, LSTM, Mobilenet etc. and implements optimization techniques such as PSO, GA, Jelly Fish optimizer etc. to extract the optimal feature and also incorporating experimental data for validation, thereby further improving epitope prediction accuracy and expanding the model's applicability to other viruses and organisms.

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